

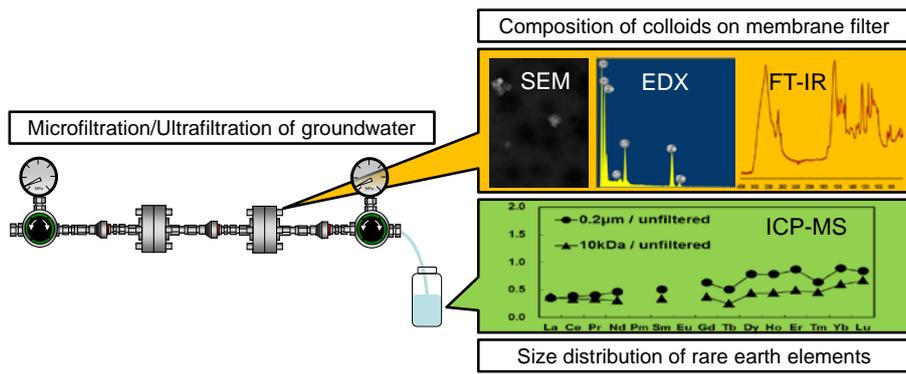


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Title	Size and composition analyses of colloids in deep granitic groundwater using microfiltration/ultrafiltration while maintaining in situ hydrochemical conditions
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Highlights

- Filtration technique while maintaining hydrochemical conditions was developed.
- Applicability of the filtration technique for analyzing colloids was demonstrated.
- Complex formation of colloids with rare earth elements was suggested.



Graphical abstract

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Size and composition analyses of colloids in deep granitic groundwater using microfiltration/ultrafiltration while maintaining in situ hydrochemical conditions

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13 **Abstract**

14 In studies of colloids in deep groundwater, a serious problem exists because the properties of the
15 colloids are influenced by changes in the chemistry of groundwater upon exposure to the
16 atmosphere and pressure release during sampling. To collect and analyze colloids in deep
17 groundwater without significantly changing the groundwater chemistry, a
18 microfiltration/ultrafiltration technique, while maintaining in situ hydrochemical conditions
19 (mainly hydraulic pressure and anaerobic condition), was developed. The size distributions and
20 compositions of colloids in granitic groundwater obtained at a depth of 300 m at the Mizunami
21 Underground Research Laboratory (MIU) in Japan were studied using the
22 microfiltration/ultrafiltration technique. The shapes and compositions of the fractionated colloids
23 collected on the membrane filter were determined by scanning electron microscopy,
24 energy-dispersive X-ray spectroscopy, and attenuated total reflection infrared spectroscopy.
25 Additionally, partitioning of rare earth elements (REEs) based on colloid size was compared with
26 REE concentrations in the filtered and unfiltered groundwater samples using inductively coupled
27 plasma mass spectrometry. Different types of colloids consisting of inorganic and organic
28 substances were observed and complex formation between the colloids and REEs was suggested.
29 Furthermore, characteristic partitioning of REEs depending on colloids size was observed in
30 complex natural environments without chemical disturbance. The current findings are useful for
31 understanding migration of radionuclides.

32

33 **Keywords**

34 Ultrafiltration, Microfiltration, Groundwater, Colloids, Rare earth elements

35

36 **1. Introduction**

37 Understanding the behavior of elements in underground environments is important for
38 geological disposal of high-level radioactive waste (HLW), carbon dioxide capture and storage,
39 and groundwater pollution. In particular, the migration of radionuclides is of great concern in the
40 geological disposal of HLW. The behavior of elements in underground environments is mainly
41 controlled by groundwater flow, precipitation, diffusion, and interaction with rocks (e.g.,
42 adsorption and filtration). Additionally, it has been clearly shown that the migration velocity of
43 elements either increases or decreases in the presence of colloids (particles and macromolecules
44 in the size range from 1 to 1000 nm) [1–4]. Therefore, studying the physicochemical properties
45 (e.g., concentration, size, shape, and chemical composition) of colloids is important for
46 understanding the behavior of elements in underground environments.

47 Colloids in deep groundwater have been studied in several papers. Degueudre et al. [5]
48 **discussed the properties of colloids from various geological formations**, ranging from crystalline
49 to sedimentary, from organic-rich to organic-poor systems, and from subsurface to very deep
50 groundwater. The authors summarized the influence of geochemical conditions, such as pH,
51 redox potential, and concentration of primary ions, on colloid properties. Geckeis et al. [6]
52 reported the size distribution of smectitic colloids and the interaction between humic colloids in
53 groundwater and trace metal ions, such as U, Th, and rare earth elements (REEs), using
54 flow-field flow fractionation (FI-FFF) combined with online inductively coupled plasma mass
55 spectrometry (ICP–MS). However, there are difficulties in collecting and analyzing colloids in
56 terms of their size, composition, and interaction with trace elements in groundwater because of
57 the low concentrations of colloids in groundwater [7, 8]. To address this issue, enrichment and
58 fractionation techniques based on colloid size have been developed. Saito et al. [9] reported a

59 colloid enrichment technique, first using ultrafiltration, and then using the focusing technique
60 with a large injection loop and the slot flow technique inherent to FI-FFF. However, the most
61 serious problem is that the physicochemical properties of colloids are likely to change owing to
62 variations in the physicochemical parameters of groundwater, such as pH, redox potential, and
63 hydraulic pressure, and exposure to the atmosphere during sampling [10]. For example, CO₂
64 degassing of groundwater resulting from a decrease in the hydraulic pressure may cause an
65 increase in the pH. Precipitation of metal (hydr)oxides can occur upon oxidation of groundwater.
66 These changes influence the properties of the colloids in groundwater. Thus, a method for
67 collecting and analyzing colloids while maintaining in situ hydrochemical conditions (mainly
68 hydraulic pressure and anaerobic condition) without chemical disturbance is required. Moreover,
69 understanding the interaction between colloids and REEs in natural environments is important;
70 REEs can be regarded as analogues of trivalent actinides [11]. The hydrogeochemistry of REEs
71 in groundwater has been studied in terms of their usefulness as tracers for various geochemical
72 processes such as water–rock interactions and hydrothermal activity. REEs are known to have
73 high affinities for colloids owing to their large ionic potential (ionic charge/ionic radius) [12–21].
74 However, the interaction between colloids and REEs is not well understood. Interactions
75 evaluated on a laboratory scale do not necessarily correspond to that measured in in situ
76 experiments possibly because of chemical disturbance. For example, preferential adsorption of
77 heavy REEs (HREEs) onto iron (hydr)oxide colloids has been determined from laboratory
78 experiments [22], whereas preferential adsorption of light REEs (LREEs) onto colloids has been
79 reported in in situ experiments [23–25].

80 The objectives of this study are to develop a microfiltration/ultrafiltration technique while
81 maintaining in situ hydraulic pressure and anaerobic condition and to understand the partitioning

82 of REEs in the presence of colloids in complex natural environment without chemical
83 disturbance. Using the technique developed herein, we sampled colloids and filtered
84 groundwater in granite at a depth of 300 m that corresponds to the depth of geological disposal
85 sites for HLW according to the Designated Radioactive Waste Final Disposal Act [26]. Moreover,
86 to understand the influence of hydrochemical conditions (mainly hydraulic pressure and
87 anaerobic condition) on colloid properties, the groundwater at a depth of 200 m was exposed to
88 the atmosphere for different periods of time and fractionated by microfiltration/ultrafiltration.
89 This study provides useful information on the size distribution and composition of colloids, and
90 the interaction between colloids and REEs in complex natural environments.

91

92 **2. Development of the microfiltration/ultrafiltration apparatus**

93 The microfiltration/ultrafiltration apparatus developed in this study consisted of two filter
94 holders: an inlet pressure regulator and an outlet pressure regulator (Fig. 1). The filter holders
95 were made of stainless used steel (SUS) and designed to maintain anaerobic conditions under
96 high hydraulic pressures (10 MPa). To prevent contamination from SUS, a part of the apparatus
97 that had contact with groundwater was passivated. The concentration of Fe in ultrapure water
98 flowing through the apparatus was less than 0.01 µg/L and that of REEs was less than 0.1 ng/L.
99 The results confirmed that colloid characterization was not influenced by contamination from
100 SUS. Microfiltration/ultrafiltration membranes with different pore sizes were packed into the
101 filter holders. For example, when membranes with pore sizes of 0.45 and 0.01 µm were set into
102 the pre- and post-filter holders, respectively, colloids in the size range of 0.01–0.45 µm were
103 collected. The inlet and outlet pressure regulator valves connected to the filter holders were used
104 to adjust the differential pressure to prevent excess resistance pressure (~0.4 MPa) of the

105 microfiltration/ultrafiltration membrane filter in the filter holder. Thus, though the pressure of
106 groundwater is higher than the resistance pressure of the membrane filter,
107 microfiltration/ultrafiltration can be performed without failure of the membrane filter. The inlet
108 pressure regulator, outlet pressure regulator, and two filter holders were connected using
109 quick-connects (Swagelok, Solon, OH, USA) with double-end shut-off stems to prevent
110 exposure to the atmosphere. Though the filter holders are separated from the inlet and outlet
111 pressure regulators after microfiltration/ultrafiltration, **hydraulic pressure and anaerobic**
112 **condition** in the filter holder can be maintained. Following discharge of residual groundwater in
113 the filter holder using inert gas (e.g., Ar), the membrane filters were collected from the filter
114 holder in an anaerobic chamber purged with an inert gas. The relatively small package size (180
115 mm × 500 mm × 120 mm; 3.2 kg) of the microfiltration/ultrafiltration apparatus affords easy
116 transport and assembly in underground facilities. Moreover, the present
117 microfiltration/ultrafiltration technique can be conducted under artesian pressure as the driving
118 force without pumping that may cause artificial alteration.

119

120 **3. Materials and methods**

121 *3.1. Sampling point*

122 Groundwater samples were collected from boreholes 07MI07 and 09MI20. **07MI07 and**
123 **09MI20 boreholes are both horizontal boreholes with the lengths of 55 and 102 m, respectively,**
124 **located on the -200 and -300 m access/research galleries of MIU, respectively.** The geological
125 site at MIU consists of Cretaceous granitic rocks (Toki granite; ~60–70 Ma) and Miocene
126 sedimentary rocks (Mizunami Group; ~15–20 Ma). The Mizunami Group unconformably
127 overlies the Toki granite at a depth of ~160 m at the MIU site. The 07MI07 and 09MI20

128 boreholes are designed for investigations of any hydrochemical changes related to facility
129 construction. The 07MI07 and 09MI20 boreholes are divided into six sections by impermeable
130 packers. The sections are numbered from 1 to 6 according to the distance from the base of the
131 07MI07 and 09MI20 boreholes. Groundwater samples were collected from each section through
132 nylon tubing without exposure to the atmosphere. The outlet of the nylon tubing was directly
133 connected to the microfiltration/ultrafiltration apparatus developed in this study using
134 quick-connects with a double-end shut-off stem, i.e., a valve was used to simplify connection and
135 disconnection of the microfiltration/ultrafiltration apparatus from the nylon tubing.

136

137 *3.2. Air exposure experiment*

138 Microfiltration of groundwater samples collected from section 4 of borehole 07MI07 at a
139 depth of 200 m was performed under different exposure times to the atmosphere (0, 180, and
140 43,200 min) following pressure release. The sample exposed to the atmosphere for 0 min was
141 filtered using the apparatus mentioned above under artesian pressure (~1 MPa). A microfiltration
142 membrane with a pore size of 0.2 μm (H020A047A, Advantec Toyo Kaisha, Ltd., Tokyo, Japan)
143 was used. The samples exposed to the atmosphere for 180 and 43,200 min were filtered using a
144 filter holder (SAKE-142, Advantec Toyo Kaisha, Ltd., Tokyo, Japan) in the laboratory under gas
145 pressure (~0.3 MPa) after exposure to atmosphere and pressure release in a Teflon bottle. A
146 microfiltration membrane with a pore size of 0.2 μm (H020A142C, Advantec Toyo Kaisha, Ltd.,
147 Tokyo, Japan) was used.

148

149 *3.3. Microfiltration/ultrafiltration under maintained in situ conditions*

150 Microfiltration/ultrafiltration of groundwater samples collected from sections 1–6 of borehole

151 09MI20 at a depth of 300 m was performed using the apparatus mentioned above under artesian
152 pressure. To remove large particles, membrane filters with a pore size of 0.45 μm (A045A047A,
153 Advantec Toyo Kaisha, Ltd., Tokyo, Japan) were fitted into pre-filter holders. A microfiltration
154 membrane with a pore size of 0.2 μm (H020A047A, Advantec Toyo Kaisha, Ltd., Tokyo, Japan)
155 or an ultrafiltration membrane with a molecular weight cut-off (MWCO) of 10 kDa (13622,
156 Merck, Darmstadt, Germany) was used for the post-filter. The filtered volume was 1 L for both
157 types of filtration. The duration of the micro- and ultrafiltration was ~0.5 and 10 h, respectively.
158 The in situ groundwater pressure as displayed on the inlet pressure gauge remained mostly
159 constant and was comparable with the in situ pressure (~2 MPa) during filtration. The differential
160 pressure between the inlet and outlet pressures was adjusted to ~0.3 MPa at the start of filtration,
161 and remained mostly constant during filtration. The dissolved oxygen (DO) was measured by
162 colorimetric method. The DO concentration in the unfiltered groundwater was less than 0.02
163 mg/L, however, rapidly increased upon exposure to the atmosphere. The DO concentration of
164 groundwater filtered using the microfiltration/ultrafiltration apparatus was less than 0.02 mg/L
165 even after sample collection and differed slightly from that of unfiltered groundwater. This
166 indicates that the **hydraulic pressure and anaerobic condition** of groundwater are maintained
167 during the filtration process.

168

169 *3.4. Characterization of colloids*

170 *3.4.1. ICP–MS measurements*

171 The concentrations of Fe and REE in the filtered and unfiltered groundwater were determined
172 by ICP–MS (ELAN DRC-II, PerkinElmer Inc., Waltham, MA, USA). The REE concentrations
173 were determined after preconcentration using chelate resin (Chelate Disc Empore, Sumitomo 3M

174 Ltd., Tokyo, Japan). The calibration curves were constructed from 0.01, 0.05, 0.1, 0.5, 1, 5, and
175 10 µg/L REE solutions prepared from 10 mg/L REE standard solution (SPEX CertiPrep Ltd.
176 London, UK). Sample preparation and ICP–MS measurements were conducted in a clean room
177 (Class 10,000). The detection limit values of Fe and REEs were 0.01 µg/L and 0.1 ng/L,
178 respectively.

179

180 *3.4.2. Scanning electron microscopy–energy-dispersive X-ray spectroscopy*

181 After filtration, the 10 kDa MWCO membrane filters were dried in an anaerobic chamber
182 under Ar atmosphere. The dry membrane filters were coated with Os. The filters were imaged
183 using a field-emission scanning electron microscope (FE–SEM, Quanta 200, FEI Co., Hillsboro,
184 OR, USA) equipped with an energy-dispersive X-ray analyzer (EDX, INCA Energy version 4.05,
185 Oxford Instruments plc., Abingdon, UK) under high vacuum conditions. Sample preparation was
186 performed in a clean room (Class 10,000). The accelerating voltage was 15 kV. The
187 magnification was in the range of 100–40,000×. The equivalent circle diameters of the colloids
188 were calculated by image processing of the obtained backscattered electron images.

189

190 *3.4.3. Fourier transform infrared spectroscopy*

191 After filtration, the 10 kDa MWCO membrane filters were dried in an anaerobic chamber
192 under Ar atmosphere. Each filter was examined by attenuated total reflection infrared
193 spectroscopy (ATR-FTIR). The ATR-IR spectra (650-4000 cm⁻¹) were recorded on a Fourier
194 transform infrared spectrophotometer (Spectrum Spotlight400, PerkinElmer Inc.).

195

196 **4. Results and discussion**

197 *4.1. Air exposure experiment*

198 To understand the influence of the hydrochemical conditions (**mainly hydraulic pressure and**
199 **anaerobic condition**) on the colloid properties, filtrations were performed under different
200 exposure times to the atmosphere (0,180, and 43,200 min) following pressure release.

201 **Table 1** shows the concentration of Fe in the filtered groundwater for different exposure times
202 to the atmosphere. The Fe concentration in the filtered groundwater sample at 180 min was lower
203 than that at 0 min. This indicates that Fe colloids larger than 0.2 μm formed by oxidation within
204 180 min after sample collection. The Fe concentrations at 180 and 43,200 min were not markedly
205 different, suggesting that the formation of Fe colloids may be a rapid process reaching
206 equilibrium quickly. The results of the Fe concentration indicate that exposure to the atmosphere
207 during filtration impedes on the concentration determination of redox-sensitive elements in the
208 filtered groundwater sample. For instance, Fe colloid formation as a result of oxidation may lead
209 to an overestimation of the colloid size.

210 **Fig. 2** shows the relative REE concentrations in the filtered groundwater samples for different
211 exposure times to the atmosphere (180 and 43,200 min) after pressure release. Each
212 concentration was normalized to filtered groundwater at 0 min. The data at 180 and 0 min were
213 not markedly different, suggesting that REEs remained in the $<0.2 \mu\text{m}$ fraction despite formation
214 of Fe colloids. In contrast to the data at 180 min, the REE concentrations in the filtered
215 groundwater sample at 43,200 min were less than those at 0 min, especially for LREEs (La, Ce,
216 Pr, and Nd). This indicates that REE adsorption onto the Fe colloids occurred within 180–43,200
217 min; however, Fe colloids were formed within 180 min. **This discrepancy** may be due to the
218 degassing of carbon dioxide from groundwater after pressure release. Carbonate ions can form
219 stable complexes with REEs in groundwater, especially with heavier REEs [27] under both

220 pressurized and anaerobic conditions. Most HREEs–carbonate complexes ($<0.2 \mu\text{m}$) were stable
221 after 43,200 min. On the other hand, a part of LREE-carbonate complexes ($<0.2 \mu\text{m}$) might have
222 dissociated within 180–43,200 min upon degassing of carbon dioxide owing to exposure to the
223 atmosphere and pressure release because the stability of LREE-carbonate complexes are weaker
224 than that of HREEs–carbonate complexes. Then, the free ion of LREEs may be adsorbed onto Fe
225 colloids ($>0.2 \mu\text{m}$). However, this hypothesis was not verified because the concentration of
226 inorganic carbon and pH were not measured. Based on these results, the oxidation of
227 groundwater influenced the size distribution of the REEs though the REEs (except for Ce and
228 Eu) were insensitive to the redox conditions.

229

230 4.2. Size distribution of colloids and REE partitioning

231 The REE concentrations in the unfiltered groundwater of the 09MI20 borehole are shown in
232 Fig. 3. The groundwater samples collected from all sections contained 0–25 ng/L REEs. The
233 rock–water interaction is the main source of REEs because granitic rocks contain relatively large
234 amounts of REE [28]. The difference in the REE concentrations between the sections suggests
235 spatial variation of REEs. Mizuno et al. [29] reported that the salinity of groundwater changes
236 according to variations in the flow conditions of groundwater owing to the shaft construction at
237 MIU. Therefore, the spatial variation of REEs may result from the shaft construction as well as
238 the salinity. Moreover, a short sampling time is required to understand the size distribution of
239 colloids and REEs in groundwater during construction of the facilities.

240 Fig. 4 shows the relative REE concentrations in the filtered groundwater samples collected
241 from the six sections of 09MI20 borehole. The groundwater samples were filtered using
242 membranes with different pore sizes (10 kDa MWCO and $0.2 \mu\text{m}$). Each concentration was

243 normalized to the unfiltered groundwater. All samples were collected on July 21, 2011 and July
244 26, 2011. For samples collected from section 2, the REE concentrations in the filtered
245 groundwater samples were higher than those of the unfiltered groundwater. This was believed to
246 be due to an analytical error during ICP–MS measurements because the REE concentrations of
247 samples from section 2 were the lowest among samples collected from all sections of 09MI20
248 borehole and comparable with the detection limit of the ICP–MS instrument. Thus, results
249 relating to samples from section 2 are not discussed. The REE concentrations in the filtered
250 groundwater decreased with decreasing membrane pore sizes (except for sample collected from
251 section 2). The results indicate REE association with colloids present with size ranges of 10
252 kDa–0.2 μm and 0.2–0.45 μm . About 50% of the REEs present in all sections (except for section
253 2) associated with colloids with sizes larger than 10 kDa. Preferential association of LREEs with
254 colloids with sizes ranging from 10 kDa to 0.2 μm and from 0.2 to 0.45 μm was observed in
255 samples collected from sections 5 and 6, whereas that of LREEs with colloids with sizes ranging
256 from 0.2 to 0.45 μm was observed in samples collected from section 1. Preferential association
257 of LREEs with colloids, as observed in groundwater, has also been observed in river water [30].

258

259 *4.3. Size distribution and composition of colloids on the ultrafiltration membrane*

260 **Fig. 5** shows representative SEM images of the inorganic colloids in the groundwater samples
261 collected from all six sections of 09MI20 borehole. The equivalent circle diameters of the
262 inorganic colloids were estimated by image processing of backscattered electron images. For all
263 sections, the equivalent circle diameters of most of the inorganic colloids were in the range of
264 0.05–0.45 μm . These results were consistent with the size distribution of the colloids estimated
265 from the size distribution of REEs, as mentioned in section 4.2. In this study, it was difficult to

266 measure the equivalent circle diameters of colloids smaller than 0.05 μm . High-resolution
267 analysis techniques, such as transmission electron microscopy and atomic force microscopy, are
268 required for such measurements.

269 Fig. 6 (a) shows representative SEM images of the inorganic colloids in the groundwater
270 samples collected from all six sections of the 09MI20 borehole and Fig. 6 (b) shows EDX
271 spectra of the inorganic colloids shown in Fig. 6 (a). The spectra show the presence of C, O, and
272 S that are from the polysulfone membrane filter. Inorganic particles containing Fe believed to be
273 iron sulfide and iron (hydr)oxide were found in samples collected from sections 1 and 3.
274 Inorganic particles containing Al, Si, alkaline earths, and alkaline earth metals were found in
275 samples collected from sections 2, 5, and 6. Inorganic particles containing Al and Mg were found
276 in samples collected from section 4. These particles are believed to be fragments of either granite
277 or clay minerals. Because Fe (hydr)oxide and clay minerals have high affinities for REEs
278 [14–18], inorganic colloids seem to form complexes with the REEs.

279 Vilks et al. [31] reported that organic colloids with a range of sizes were present together with
280 inorganic colloids. Thus, the effect of organic colloids on partitioning of REEs was considered.
281 Identification of organic matter by SEM was difficult because the membrane filter was made of
282 an organic substance. To confirm the presence of organic matter, IR analysis was performed. The
283 FTIR spectra of all six sections and the Suwannee River humic acid standard (2S101H,
284 International Humic Substances Society) are shown in Fig. 7. All FTIR spectra were
285 characterized by a number of absorption bands, exhibiting varying relative intensities. The strong
286 broad band at 2400–3700 cm^{-1} is generally attributed to O–H stretching of hydroxyl groups
287 involved in hydrogen links, carboxyl groups, and phenol groups. The band at 3292 cm^{-1} , which
288 was weak in the humic acid spectrum, was attributed to the N–H stretch absorption band of

289 amines and amides [32,33]. There were also a few absorption bands at 2923 and 2853 cm^{-1} .
290 These bands were assigned to C–H stretching of methyl (CH_3) and methylene (CH_2) groups of
291 aliphatic chains [34–36]. The bands at 1730 and 1701 cm^{-1} are generally attributed to C=O
292 stretching vibrations, mainly of carboxyl groups and, to a lesser extent, of ketones and aldehydes
293 [34–36]. The band at 1644 cm^{-1} , which was absent from the humic acid spectrum, was ascribed
294 to C=C stretching of aromatic rings [37] and C=O stretching vibration in amides (amide I band)
295 [38]. The band at 1605 cm^{-1} , which was absent from the spectra in all groundwater samples, was
296 ascribed to stretching of the C=C group conjugated with another C=C group (aromatic ring),
297 C=O and/or COO^- in unsaturated ketones, carboxylic acids, and amides [39]. The band at 1545
298 cm^{-1} , which was absent from the humic acid spectrum, was ascribed to NH bending in secondary
299 amides (amide II band) [38]. All samples exhibit broad bands in the region 1000–1300 cm^{-1} ,
300 which are probably because of the C–O stretching vibration in alcohols, ethers, phenols,
301 carboxylic acids, and esters [38, 40]. For the groundwater samples collected from all sections of
302 the borehole, it was difficult to detect absorption bands in the region of 650–1400 cm^{-1} because
303 of the strong absorption bands of the blank.

304 The FTIR spectra of organic matter present in samples collected from all sections showed
305 similar shapes and intensities, suggesting that all sections of the 09MI20 borehole were similar
306 structures. Some differences were observed between the FTIR spectra of organic matter and
307 humic acid. Although the organic matter showed some absorption bands relating to nitrogen,
308 such as absorption bands at 1545, 1644, and 3292 cm^{-1} , these absorption bands were weak or
309 different from those of humic acid. These differences indicated that the nitrogen content of the
310 organic matter in the groundwater samples collected from all sections of the borehole was greater
311 than that of humic acid. Previous studies reported that most of the REEs in groundwater bound to

312 humic substances by forming stable complexes [41]. Organic matter present in 09MI20 borehole
313 may associate with REEs as well as inorganic colloids.

314 Vilks et al. [31] reported that inorganic colloids consisting of clay minerals, ferrioxide, and
315 carbonate minerals were present in a wide size range with organic matter in groundwater. Saito et
316 al. [9] reported that Al-, Mg-, and Fe-bearing colloids associated with organic matter at a depth
317 of 300 m at the MIU site. Additionally, in this study, we reveal that organic matter in
318 groundwater associated with inorganic colloids and clarified the structures of organic matter.
319 Although the results obtained from this study are qualitatively similar to those of previous
320 studies [9, 31], our results may give more useful information owing to the newly developed
321 sampling technique that can be operated under in situ hydrochemical conditions.

322

323 5. Conclusions

324 In this study, we developed a microfiltration/ultrafiltration technique that can maintain in situ
325 physicochemical parameters and afford collection of colloids without chemical disturbance. The
326 microfiltration/ultrafiltration technique can solve some of the uncertainties regarding colloid
327 properties such as size distribution and interaction with REEs.

328 Using the microfiltration/ultrafiltration technique developed in this study, we obtained useful
329 information on the presence of inorganic (e.g., Fe, Al, Mg, and Si) and organic colloids with a
330 wide size range, and REEs characteristic partitioning in granite groundwater at a depth of 300 m.
331 The latter depth corresponds to that of geological disposal sites for high-level radioactive waste.
332 The inorganic colloids are primary minerals (i.e. fragments of granites) or secondary minerals
333 such as clay minerals resulting from dissolution and precipitation of the host rock. Moreover, the
334 findings indicated the co-existence of inorganic and organic colloids, and colloids bound metal

335 ions. REEs characteristic partitioning, such as preferential association of LREEs with colloids
336 with sizes ranging from 10 kDa to 0.2 μm and from 0.2 to 0.45 μm in a specific section of
337 borehole, may be due to the influence of the wide variety of colloids. In future studies, we will
338 investigate the effect of these colloids on radioactive behaviors using combined
339 microfiltration/ultrafiltration technique and bulk analyses.

340

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345

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455

456 **Figure captions**

457 **Fig. 1.** Schematic illustration of the microfiltration/ultrafiltration apparatus operating
458 under pressurized/anaerobic conditions.

459 **Fig. 2.** Relative REE concentrations in the filtered groundwater samples at different
460 exposure times to the atmosphere (180 and 43,200 min). Each concentration was
461 normalized to filtered groundwater sample at 0 min.

462 **Fig. 3.** Concentrations of REEs in the groundwater samples collected from all six
463 sections of the 09MI20 borehole.

464 **Fig. 4.** Relative REE concentrations in the filtered groundwater samples collected from
465 all six sections of the 09MI20 borehole. The groundwater samples were filtered
466 using membranes with different pore sizes (10 kDa MWCO and 0.2 μm). Each
467 concentration was normalized to the unfiltered groundwater sample.

468 **Fig. 5.** Representative SEM images of inorganic colloids in the groundwater samples
469 collected from all six sections of the 09MI20 borehole. The equivalent circle
470 diameters of several colloids are shown.

471 **Fig. 6.** Representative SEM images and EDX spectra of inorganic colloids in the
472 groundwater samples collected from all six sections of the 09MI20 borehole: (a)
473 SEM images and (b) EDX spectra of the inorganic colloids shown in Fig. 6 (a).

474 **Fig. 7.** FTIR spectra of the reference humic acid and organic colloids in the
475 groundwater samples collected from all six sections of the 09MI20 borehole.

476

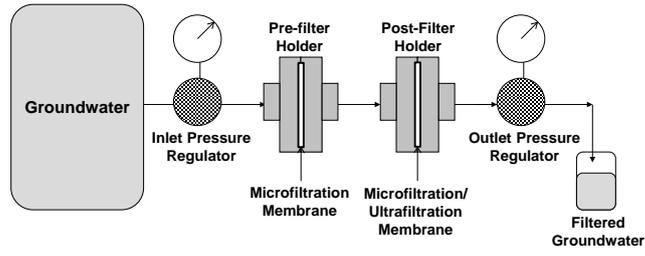


Fig. 1. Schematic illustration of the microfiltration/ultrafiltration apparatus operating under pressurized/anaerobic conditions.

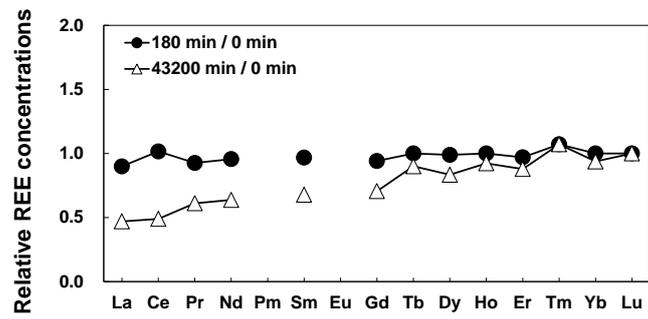


Fig. 2. Relative REE concentrations in the filtered groundwater samples at different exposure times to the atmosphere (180 and 43,200 min). Each concentration was normalized to filtered groundwater sample at 0 min.

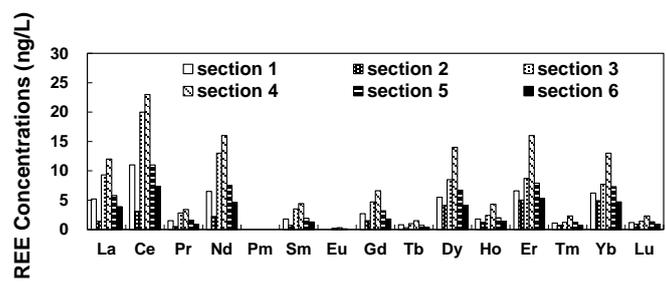


Fig. 3. Concentrations of REEs in the groundwater samples collected from all six sections of the 09MI20 borehole.

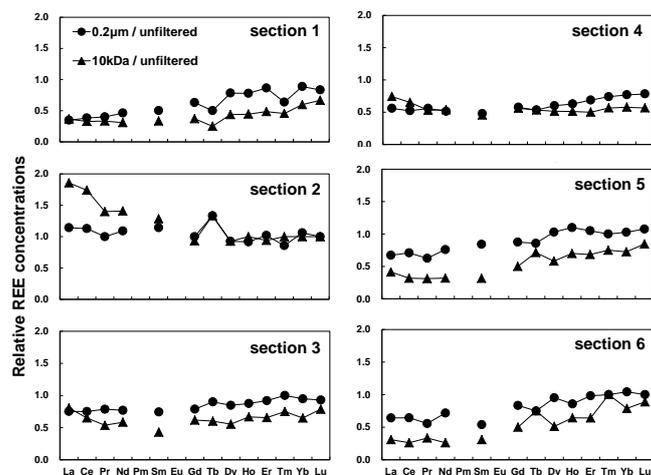


Fig. 4. Relative REE concentrations in the filtered groundwater samples collected from all six sections of the 09MI20 borehole. The groundwater samples were filtered using membranes with different pore sizes (10 kDa MWCO and 0.2 μm). Each concentration was normalized to the unfiltered groundwater sample.

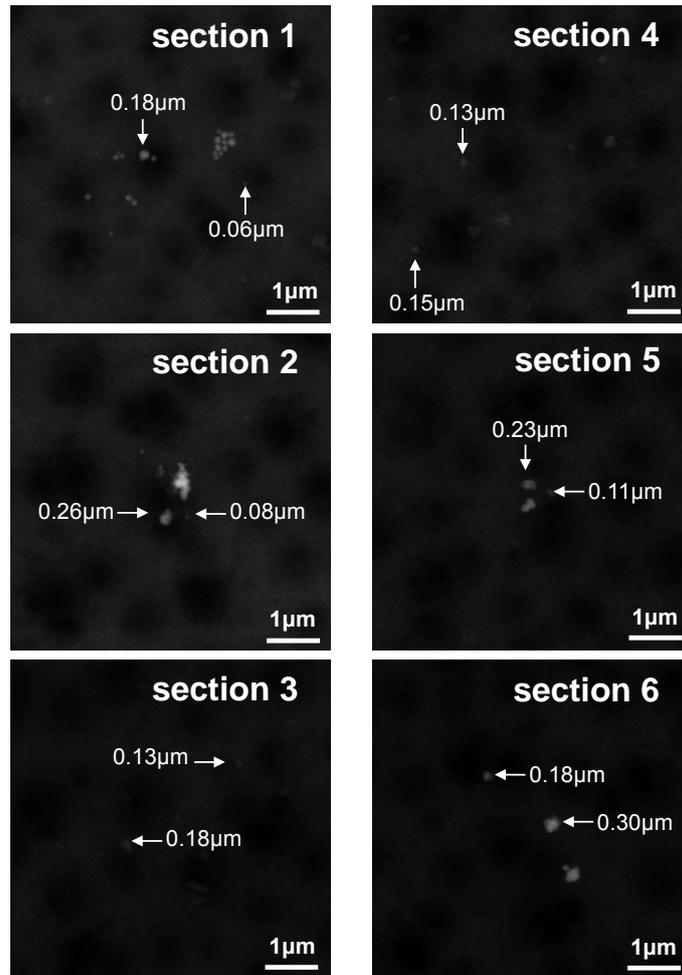


Fig. 5. Representative SEM images of inorganic colloids in the groundwater samples collected from all six sections of the 09MI20 borehole. The equivalent circle diameters of several colloids are shown.

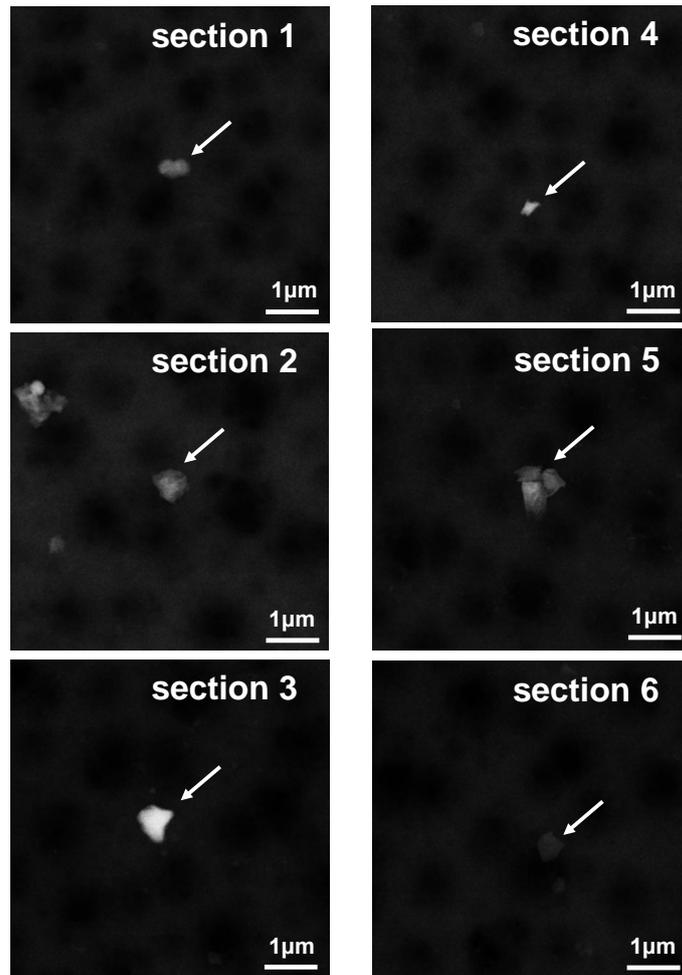


Fig. 6(a). Representative SEM images and EDX spectra of inorganic colloids in the groundwater samples collected from all six sections of the 09MI20 borehole: (a) SEM images and (b) EDX spectra of the inorganic colloids shown in Fig. 6 (a).

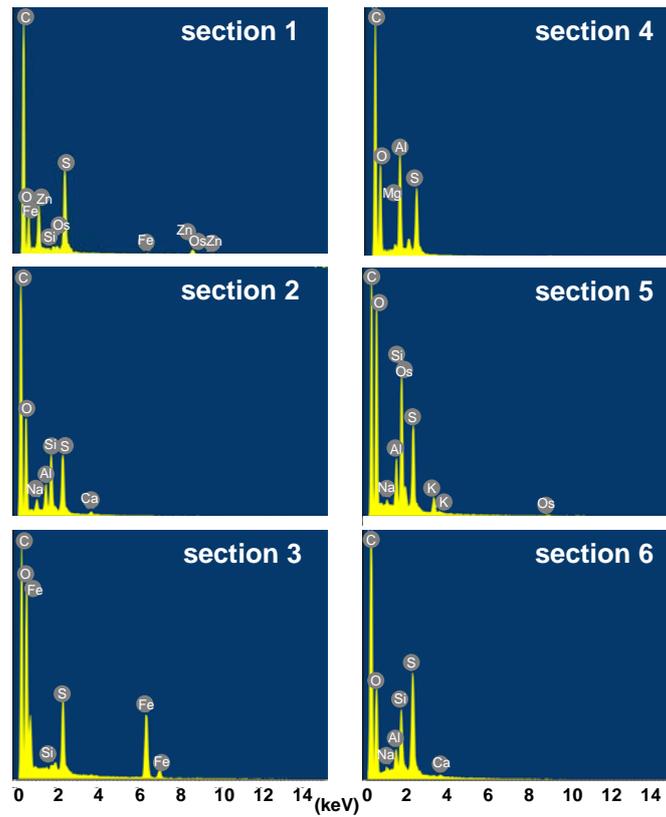


Fig. 6(b). Representative SEM images and EDX spectra of inorganic colloids in the groundwater samples collected from all six sections of the 09MI20 borehole: (a) SEM images and (b) EDX spectra of the inorganic colloids shown in Fig. 6 (a).

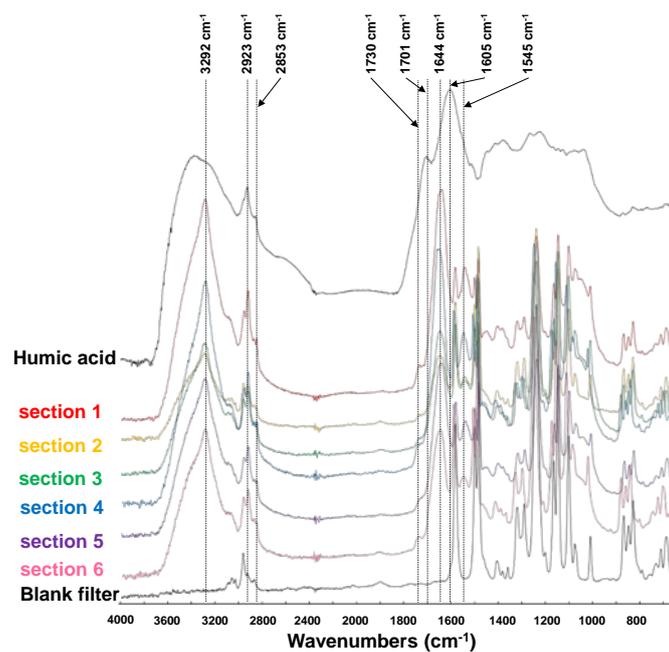


Fig. 7. FTIR spectra of the reference humic acid and organic colloids in the groundwater samples collected from all six sections of the 09MI20 borehole. FTIR spectra of the reference humic acid and organic colloids in the groundwater samples collected from all six sections of the 09MI20 borehole.

Table 1

Average concentrations of total Fe in filtered groundwater.

	Exposure time					
	0 (min)		180 (min)		43200 (min)	
	Avg.	S.D ^a	Avg.	S.D ^a	Avg.	S.D ^a
Fe (µg/L)	5.5	0.52	3.2	0.42	2.7	0.28

^aThe standard deviation of three samples.