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Efficient condensation of organic colloids in deep groundwater using surface-modified nanofiltration membranes under optimized hydrodynamic conditions

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14 **Abstract**

15 The transport of radionuclides by organic colloids in deep groundwater is one of the important
16 issues for the geological disposal of high-level radioactive waste. Because of their low
17 concentration, it is difficult to directly analyze organic colloids in deep groundwater. Hence, it is
18 useful to utilize porous membranes for the condensation which can increase the concentration of
19 organic colloids in groundwater physically without alteration of their properties, although some
20 part of the components is lost by membrane fouling. In this study, hydrodynamic conditions were
21 optimized, and surfaces of nanofiltration (NF) membranes were modified using a cationic
22 phosphorylcholine polymer, poly(2-methacryloyloxyethyl phosphorylcholine-co-2-aminoethyl
23 methacrylate) (p(MPC-co-AEMA)), for preventing membrane fouling and improving
24 condensation efficiency. Aqueous solutions of humic acid or bovine serum albumin (BSA) were
25 used as models of organic colloids, and they were condensed using a laboratory-scale cross-flow
26 filtration apparatus equipped with a commercial sulfonated polyethersulfone NF membrane. The
27 effects of hydrodynamic conditions, such as the applied transmembrane pressure (TMP) and
28 stirring rate, and membrane surface modification on condensation efficiency were evaluated. A
29 low TMP and high stirring rate effectively improved the recovery yield of humic acids and BSA.
30 The membrane surface coated with p(MPC-co-AEMA) was significantly effective for preventing
31 the decline in permeate flux, caused by fouling with BSA. Deep groundwater, obtained from a
32 depth of 300 m at the Mizunami Underground Research Laboratory in Japan, was condensed.
33 The recovery yield of the organic colloids in the deep groundwater condensation test at 5-fold
34 condensation was improved from 62% to 92% by the optimized hydrodynamic conditions and
35 membrane surface modification for prevention of membrane fouling. The composition of organic
36 colloids in the condensates was analyzed using pyrolysis gas chromatography coupled with mass

37 spectrometry.

38

39 Keywords: Condensation, Nanofiltration membrane, Organic colloid, Groundwater,

40 Hydrodynamic condition, Membrane surface modification

41 **1. Introduction**

42 High-level radioactive waste (HLW) originating from the nuclear industry is legislated to be
43 disposed of underground at a depth of more than 300 m [1]. The safety of the geological disposal
44 of HLW is dependent on the migration of the radionuclides released from HLW into the
45 underground environment. Organic colloids are present in deep groundwater [2,3], and the
46 interaction between organic colloids and radionuclides affects the migration of radionuclides
47 [4,5]. Therefore, for the safety assessment of HLW, it is imperative to understand the detailed
48 composition of organic colloids and their interaction with radionuclides. Precise analysis of
49 organic colloids from groundwater is difficult, caused by their low concentration [6,7]; hence,
50 extraction techniques using adsorption resins [8,9] are widely applied. However, extracted
51 samples using adsorption resins are exposed to severe chemical disturbances, and there is
52 concern over their chemical changes. On the other hand, condensation using porous membranes
53 has also been applied to afford high concentrations of organic colloids. Although porous
54 membranes can rapidly condense organic colloids without chemical exposure [10–12], some
55 amount of organic colloids from the condensed samples is lost because of their deposition on the
56 membrane surface and subsequent blocking of pores (membrane fouling). For obtaining more
57 accurate information on organic colloids, it is imperative to prevent membrane fouling.

58 Effects of hydrodynamic conditions on membrane fouling by organic colloids have been
59 investigated [13–15]. These studies have reported that membrane fouling is affected by chemical
60 and hydrodynamic interactions. That is, the fouling of membranes by organic colloids is
61 promoted by high concentrations of divalent cations (Ca^{2+} and Mg^{2+}), high initial permeate flux,
62 and low cross-flow velocity. Divalent cations neutralize the net negative charge of humic acids,
63 which supposedly constitute the majority of organic colloids in groundwater, resulting in the

64 reduction of repulsion among humic acid molecules [16–18]. In addition, divalent cations can
65 bridge humic acid molecules [19]. Thus, divalent cations accelerate the fouling of membranes,
66 caused by the coagulation of humic acid molecules. The high permeate flux or low cross-flow
67 velocity increases the concentration of divalent cations on the membrane surface, caused by
68 concentration polarization, which in turn enhances membrane fouling [14]. Thus, increase in the
69 cross-flow velocity and decrease in the permeate flux mitigate the fouling of membranes by
70 organic colloids.

71 The surface modification of membranes using various hydrophilic polymers has also been
72 effective for preventing membrane fouling [20]. Hydrophilic polymers, such as polyethylene
73 glycol, and zwitterionic polymers have been frequently used for modifying the surfaces of
74 porous membranes with the aim of preventing the adsorption of organic colloids and fouling of
75 membranes [21–24]. We have developed a simple, facile method for modifying polyamide
76 reverse osmosis (RO) membranes using zwitterionic polymers with the aim of preventing the
77 adsorption of bacteria [25]. Negatively charged RO membranes were coated with
78 poly(2-methacryloyloxyethyl phosphorylcholine-co-2-aminoethyl methacrylate)
79 (p(MPC-co-AEMA)), which is composed of cationic AEMA units and zwitterionic MPC units,
80 by electrostatic interaction. The coated RO membranes exhibited high hydrophilicity and high
81 resistance to bacterial adsorption. Therefore, membrane surface modification with
82 p(MPC-co-AEMA) is also promising for preventing the fouling of membranes by organic
83 colloids.

84 Previously, we have developed a method for the condensation of organic colloids in
85 groundwater using nanofiltration (NF) membranes [12]. The use of NF membranes facilitated the
86 condensation of organic colloids without severe reduction in flux, while condensation using RO

87 membranes facilitated severe reduction in flux, caused by the precipitation of condensed
88 inorganic substances. Organic colloids in actual condensed groundwater at a depth of 300 m in
89 granitic rocks were successfully analyzed by pyrolysis gas chromatography coupled with mass
90 spectrometry (Py-GC/MS), and their composition was similar to that of humic substances with
91 high humification. However, the recovery yields of organic colloids were up to 57% at 20-fold
92 condensation, caused by the fouling of organic colloids. Thus, the recovery yields need to be
93 improved for the purpose of confirming the validity of the result obtained by Py-GC/MS.

94 This study aimed at preventing the fouling of membranes by organic colloids in the
95 condensation of deep groundwater using NF membranes by optimizing hydrodynamic conditions
96 and modifying the membrane surface. Commercial humic acids and bovine serum albumin
97 (BSA) were used as models of organic colloids. The aqueous solutions of organic colloids were
98 condensed using a cross-flow filtration apparatus equipped with sulfonated polyethersulfone NF
99 membranes. The time courses of permeate flux and the recovery yield of humic acid and BSA
100 were monitored. The effects of hydrodynamic conditions and membrane surface modification
101 with p(MPC-co-AEMA) on condensation behavior were examined. Finally, actual deep
102 groundwater was condensed using modified NF membranes under optimized hydrodynamic
103 conditions. The composition of the organic colloids in the condensed deep groundwater was
104 analyzed by Py-GC/MS.

105

106 **2. Experimental**

107 *2.1. Materials*

108 All solutions used in this study were prepared using ultrapure water and analytical-grade
109 chemicals. Unless specified, all chemicals were obtained from Wako Pure Chemical Industries

110 (Osaka, Japan) and were used without further purification. Commercial sulfonated
111 polyethersulfone NF membranes were obtained from Nitto Denko (NTR7450; Osaka, Japan).
112 Coal-derived humic acid (WHA; 088-04622; Wako Pure Chemical Industries) and peat-derived
113 humic acid (AHA; H16752; Sigma-Aldrich, St. Louis, MO, USA) were used after purification
114 according to a previous study [12]. A p(MPC-co-AEMA) (MPC:AEMA = 9:1, random
115 copolymer) was generously supplied by NOF Corporation (Tokyo, Japan), and its
116 weight-average molecular weight was 9.7×10^5 [25]. Coomassie brilliant blue (CBB) G-250 was
117 purchased from Polysciences Inc. (Warrington, PA, USA).

118 Groundwater in the granitic rock was collected from the 09MI20 borehole in the -300 m
119 Access/Research Gallery of Mizunami Underground Research Laboratory on December 25, 2014.
120 The borehole was divided into six sections by impermeable packers, and the sections were
121 numbered from 1 to 6 according to the distance from the base of the borehole. Groundwater
122 samples were collected from section 1 and placed in the refrigerator until experiments were
123 conducted. Detailed information of the borehole and hydrochemistry of the examined
124 groundwater has been summarized in our previous paper [12]. The groundwater contained 9.0
125 mg/L Ca^{2+} , and the pH was 8.5.

126

127 2.2. Condensation apparatus

128 Fig. 1 shows the laboratory-scale cross-flow membrane filtration apparatus used for
129 condensation experiments [12]. For condensation experiments, a feed solution was fed into a
130 cylindrical stainless steel membrane cell using a plunger pump (NPL-120; Nihon Seimitsu
131 Kagaku, Tokyo, Japan) at a constant flow rate of 9.0 mL/min. The applied transmembrane
132 pressure (TMP) was adjusted at 0.75 MPa or 1.5 MPa using a back pressure valve. The effective

133 membrane area was 8.0 cm². The permeate solution was collected in a permeate reservoir. The
134 retentate solution was circulated through the feed reservoir and was condensed. The feed solution
135 in the cell was stirred using a cylindrical magnetic bar with a length of 3.0 cm at 150 or 1500
136 rpm. Permeate flux was calculated from the time course of permeate weight.

137

138 *2.3. Modification of membrane surface using a cationic phosphorylcholine polymer*

139 A negatively charged NF membrane was modified using cationic p(MPC-co-AEMA) by
140 electrostatic interaction [25]. As shown in Fig. 1, the NF membrane was set into the membrane
141 cell. An aqueous solution of 0.1 wt% p(MPC-co-AEMA) was fed into the membrane cell at a
142 constant flow rate of 9.0 mL/min and a constant TMP of 0.75 MPa for 2 h. The feed solution of
143 the membrane cell was magnetically stirred at 150 rpm. Then, the membrane was detached and
144 washed by gentle shaking in an aqueous solution of 3.5 wt% NaCl for removing non-specifically
145 adsorbed polymers. For evaluating the surface hydrophilicity of the membranes, the water
146 contact angle was measured using a contact angle meter (DM-300; Kyowa Interface Science,
147 Saitama, Japan). The ζ -potential was measured by streaming potential measurement using an
148 electrophoretic light-scattering apparatus (ELSZ-1000, Otsuka Electronics, Osaka, Japan) in
149 aqueous 10 mmol/L NaCl.

150

151 *2.4. Condensation of humic acid and BSA*

152 A feed solution of 5 mg/L of humic acid containing 1 mg/L Ca²⁺ or 5 mg/L of BSA was
153 condensed from 125 mL to 25 mL (5-fold condensation) using the raw NF membrane and
154 p(MPC-co-AEMA)-coated NF membrane under different TMPs (0.75 or 1.5 MPa) and stirring
155 rates (150 or 1500 rpm). The concentration of Ca²⁺ in the feed solution of humic acid was

156 adjusted using a 1000 mg/L Ca^{2+} standard solution composed of CaCO_3 in 0.1 mol/L HNO_3
157 (Wako Pure Chemicals).

158 The humic acid concentration was determined by optical density measurements at 254 nm
159 using a UV–vis spectrometer (V-650; Jasco Corp., Tokyo, Japan). The BSA concentration was
160 determined by a CBB method [26]. CBB was dissolved in methanol, and then phosphoric acid
161 and ultrapure water were added. The final concentration of the components in this CBB solution
162 was 0.025% (w/v) CBB, 12.5% (w/v) methanol, and 70.83% (w/v) phosphoric acid. The CBB
163 solution (0.2 mL) and 0.8 mL of the sample were mixed, and the optical density at 595 nm was
164 measured using a UV–vis spectrometer. To evaluate condensation efficiency, the recovery yield
165 was defined as the amount of humic acid or BSA in the condensed solution divided by that in the
166 initial solution.

167

168 *2.5. Characterization of organic colloids*

169 Condensation experiments were carried out using groundwater. The concentration of organic
170 colloids in the groundwater was determined by the same method as that used for the
171 determination of humic acid under the assumption that all organic colloids are humic acids. The
172 condensed organic colloids were characterized by Py-GC/MS. Py-GC/MS analysis was carried
173 out using a double-shot pyrolyzer (PY-2020id; Frontier Laboratories Ltd., Fukushima, Japan)
174 attached to a GC/MS instrument (Agilent 6890N/Agilent 5973; Agilent Technologies Inc., Palo
175 Alto, CA, USA) Py-GC/MS is a powerful tool for obtaining detailed compositional information
176 of a polymer. Two humic acids derived from coal and peat were directly analyzed by Py-GC/MS.
177 In the case of the organic colloids in condensed groundwater, the condensed water was further
178 evaporated, dried under N_2 , and then analyzed by Py-GC/MS. Py-GC/MS analysis was

179 performed by the same method as in our previous paper [12].

180

181 **3. Results and discussion**

182 *3.1. Analysis of commercial humic acids*

183 First, two commercial humic acids WHA and AHA, which have been extensively used as
184 model humic acids by several researchers [13–15,27,28], were analyzed by Py-GC/MS. Fig. 2
185 shows the Py-GC/MS pyrogram of WHA and AHA, and Table 1 lists the detected compounds.
186 The major products obtained from the pyrolysis of coal-derived WHA were aromatic compounds
187 (n = 1, 3, 4, 5, 7, 8, 10, 13, 14, 17, 20, and 23) attributed to a high degree of humification; these
188 compounds were consistent with brown coal humic substances [29]. On the other hand, the major
189 products obtained from the pyrolysis of peat-derived AHA were a series of aliphatic compounds
190 (n = 2, 6, 8, 9, 11, 12, 15, 16, 18, 19, 21, 22, and 24–32), attributed to lipids and paraffinic
191 material in humic substances. The result obtained for AHA is similar to that obtained for
192 peat-derived humic substances [29,30]. The composition of organic colloids in deep groundwater
193 is similar to that of humic substances with high humification [12]. Thus, we used WHA as a
194 model organic colloid in the following experiments.

195

196 *3.2. Condensation of humic acid*

197 *3.2.1. Optimum hydrodynamic conditions*

198 The effects of the TMP and stirring rate on the condensation of WHA were investigated to
199 obtain optimum hydrodynamic conditions for the NF process. Fig. 3 shows the effect of the
200 hydrodynamic conditions on the permeate flux and WHA concentration in the condensed
201 solutions. In this figure, the theoretical concentrations in the condensed solutions with 100%
202 recovery yield are represented as dotted lines. The recovery yield was defined as the amount of

203 organic colloids in the condensed solution divided by that in the initial solution. Fig. 4 shows the
204 surface view of the membranes after 5-fold condensation tests of WHA. The condensation test
205 under all hydrodynamic conditions exhibited no reduction in flux (Fig. 3). However, all WHA
206 concentrations in the condensed solutions were below the dotted line, indicating low recovery
207 yields. As shown in Fig. 3a, the recovery yield after condensation at 150 rpm and 1.5 MPa was
208 quite low, only 44% (Fig. 3a). This low recovery yield was caused by the adsorption and
209 deposition of humic acid on the membrane surface. As shown in Fig. 4a, large amounts of humic
210 acids were deposited on the membrane surface. The stirring rate on the membrane surface is
211 considered greater outside the membrane cell compared with that in the center. Thus, a large
212 amount of deposits on the center of the membrane surface is probably caused by the distribution
213 of the stirring rate. However, at this time scale, these deposits hardly affected the reduction in
214 flux. As the stirring rate increased or TMP decreased, the recovery yields increased (Figs. 3b and
215 c), and the amounts of humic acids deposited on the membrane surface decreased (Figs. 4b and
216 c). The decreasing TMP and increasing stirring rate decrease the permeation drag and
217 concentration polarization of solutes on the membrane surface, resulting in the prevention of
218 membrane fouling. At a high stirring rate and low TMP, a high recovery yield of 80% was
219 obtained (Fig. 3d), with small amounts of WHA deposited on the NF membrane (Fig. 4d). These
220 results indicate that a high stirring rate and low TMP is optimum condition for high recovery
221 yield. Because the available value of the stirring rate in our apparatus was maximally limited to
222 be 1500 rpm, and a significantly low TMP led to increased experiment time, the condensation
223 test was conducted at a stirring rate of 1500 rpm and 0.75 MPa as the optimized condition. For
224 comparison, condensation tests were carried out at a stirring rate of 150 rpm and 1.5 MPa. We
225 also confirmed this optimization was effective in the case using the p(MPC-co-AEMA)-coated

226 NF membranes as described section 3.2.2 and 3.3.

227

228 *3.2.2. Modification of membrane surface*

229 To prevent the adsorption of organic colloids, the surface of the NF membrane was modified
230 with zwitterionic polymer. Fig. 5 shows the effect of the p(MPC-co-AEMA) concentration on the
231 ζ -potentials and water contact angles of the NF membranes. The ζ -potential of the NF
232 membranes was neutralized by the coating with the aqueous solution of 0.0001 wt%
233 p(MPC-co-AEMA). The water contact angle of the NF membranes decreased with increasing the
234 p(MPC-co-AEMA) concentration and reached the minimum value at 0.01 wt%
235 p(MPC-co-AEMA). Fig. 6 shows the effect of the p(MPC-co-AEMA) concentration on the
236 recovery yields and experimental times required to condensate the feed solution to 5 times on the
237 WHA condensation test. The highest recovery yield was obtained at 0.1 wt% p(MPC-co-AEMA).
238 These results indicate the coating with the aqueous solution of 0.1 wt% p(MPC-co-AEMA) made
239 the surface of the NF membranes more hydrophilic and prevented the adsorption of organic
240 colloids more effectively. Thus, the NF membranes were coated with 0.1 wt% of
241 p(MPC-co-AEMA) on the following experiments.

242 We examined the effect of the hydrodynamic conditions and membrane surface modification
243 on the recovery yields of WHA. Fig. 7 shows the time courses of the permeate flux and WHA
244 concentrations in the condensation tests using the p(MPC-co-AEMA)-coated NF membrane. Fig.
245 8 shows the surface view of the p(MPC-co-AEMA)-coated NF membrane after 5-fold
246 condensation tests of WHA. As shown in Fig. 7, in the condensation test using the
247 p(MPC-co-AEMA)-coated NF membrane (Fig. 7), reduction in flux was not observed. Although
248 the amounts of humic acids deposited on the surface of the p(MPC-co-AEMA)-coated NF

249 membrane (Figs. 8a and b) and raw NF membrane (Figs. 4a and d) seemed similar, recovery
250 yields of 56% and 86% for the p(MPC-co-AEMA)-coated NF membrane shown in Figs. 7a and b
251 were higher than those of the raw NF membrane under the same hydrodynamic condition (44%
252 and 80% shown in Figs. 3a and d, respectively). Thus, modification of the membrane surface
253 with p(MPC-co-AEMA) is effective against fouling by WHA, caused by improved hydrophilicity.
254 In addition, the optimized hydrodynamic condition was also effective to improve the recovery
255 yields in the case using the p(MPC-co-AEMA)-coated NF membrane.

256

257 *3.3. Condensation of BSA*

258 We examined the effects of hydrodynamic conditions and membrane surface modification on
259 the recovery yields of BSA. Fig. 9 shows the time courses of the permeate flux and BSA
260 concentration in the condensed solutions. The initial permeate flux value of the raw NF
261 membranes (Figs. 9a and b) was significantly lower than those obtained from the condensation
262 test of WHA under the same hydrodynamic condition (Fig. 3a and d), indicating that the
263 permeate flux of raw NF membranes drastically declines immediately after the start of the
264 condensation test of BSA. On the other hand, the permeate flux in the condensation tests using
265 the p(MPC-co-AEMA)-coated membranes (Fig. 9c and d) exhibited almost the same values as
266 those obtained from the condensation test of WHA under the same hydrodynamic conditions (Fig.
267 7a and b). Thus, the modification of the membrane surface with p(MPC-co-AEMA) is
268 significantly effective for the prevention of the decrease in permeate flux by fouling with BSA.
269 The recovery yields were significantly improved by the optimized hydrodynamic conditions (Fig.
270 9b) as compared to the membrane surface modified with p(MPC-co-AEMA) (Fig. 9c), indicating
271 that the optimized hydrodynamic conditions are more effective than the membrane surface

272 modified with p(MPC-co-AEMA) for increasing the recovery yield of BSA.

273

274 *3.4. Condensation of organic colloids in groundwater*

275 We examined the effects of hydrodynamic conditions and membrane surface modification on
276 the recovery yield of organic colloids in groundwater. Fig. 10 shows the time course of the
277 permeate flux and organic colloid concentration in the condensed solution. The condensation test
278 using the raw NF membrane under the original hydrodynamic condition exhibited no reduction
279 in flux, but the recovery yield at 5-fold condensation was only 62% (Fig. 10a). This low recovery
280 yield is attributed to the adsorption of organic colloids on the membrane surface caused by
281 membrane fouling. On the other hand, the condensation test using the p(MPC-co-AEMA)-coated
282 NF membrane under the optimized hydrodynamic condition afforded a higher recovery yield of
283 92% at 5-fold condensation (Fig. 10b). For obtaining a sufficiently high concentration of organic
284 colloids for Py-GC/MS measurement, the groundwater was condensed from 500 mL to 25 mL
285 (20-fold condensation) using the p(MPC-co-AEMA)-coated NF membrane under the optimized
286 hydrodynamic condition. Fig. 11 shows the results. The recovery yield obtained was 74% at
287 20-fold condensation, which was higher than that of 57% at 20-fold condensation in our previous
288 study using the raw NF membrane under the original hydrodynamic condition (at 150 rpm and
289 1.5 MPa) [12]. Thus, we achieved improvement in the recovery yield of organic colloids by the
290 combination of optimized hydrodynamic conditions and p(MPC-co-AEMA) coating.
291 Groundwater generally contains inorganic colloids such as fragments of granites and clay
292 minerals resulting from dissolution and precipitation of host rock [31]. Inorganic colloids would
293 also cause membrane fouling during the condensation process, although the membrane fouling
294 with inorganic colloids was not observed in this study. The optimization of hydrodynamic

295 conditions would also be promising for preventing fouling with inorganic colloids. Further
296 evaluation about the effect of inorganic colloids on condensation of organic colloids is necessary.

297

298 *3.5. Characterization of organic colloids in groundwater*

299 Fig. 12 shows the Py-GC/MS pyrogram of the organic colloids in the condensed groundwater,
300 and Table 1 lists the detected compounds. The major products obtained from the pyrolysis of
301 organic colloids were aromatic products. This result indicates that the composition of a majority
302 of the organic colloids in deep groundwater is similar to that of humic substances with high
303 humification, as has been previously reported by our group [12]. Moreover, as shown in Fig. 12b,
304 the major products obtained from the pyrolysis of p(MPC-co-AEMA), such as benzene, methyl
305 methacrylate, and dimethylethanolamine, were not detected in the pyrogram of organic colloids,
306 indicating that the p(MPC-co-AEMA) coating does not desorb from the NF membrane surface
307 and does not affect Py-GC/MS analysis. Although the result obtained from the Py-GC/MS
308 pyrogram was not different from those obtained from our previous study [12], other components,
309 which were not detected by Py-GC/MS, may be different. In particular, the result of rare earth
310 elements (REEs) could be different, caused by their high affinity with organic colloids [32–34].
311 REEs are regarded as analogs of trivalent actinides [35] and are important for HLW. Therefore,
312 REEs should be analyzed in future studies.

313

314 **4. Conclusions**

315 We examined the improvement in the recovery yields of organic colloids via the condensation
316 of deep groundwater by utilizing optimized hydrodynamic conditions, such as transmembrane
317 applied pressure and stirring rate, and modifying the surface of NF membranes with

318 p(MPC-co-AEMA). The recovery yield of humic acid from the condensation test was increased
319 by the optimized hydrodynamic conditions, attributed to the reduction of concentration
320 polarization and permeate drag, and by the modification of membrane surfaces, attributed to the
321 hydrophilization of the NF membrane surface. The recovery yield of BSA in the condensation
322 test was also increased by the optimized hydrodynamic conditions and membrane surface
323 modification. In addition, the modification of membrane surfaces was effective for preventing
324 the decrease in permeate flux, caused by BSA fouling. The recovery yield of organic colloids in
325 the condensation test of deep groundwater was significantly increased by the combination of
326 optimized hydrodynamic conditions and the membrane surface modification. The Py-GC/MS
327 analysis of organic colloids condensed by the improved method proposed in this study indicated
328 that composition of organic colloids is similar to that of humic substance with high humification,
329 same as in our previous study. The optimization of hydrodynamic conditions and modification of
330 membrane surfaces, which were effective for the improvement in the recovery of organic
331 colloids, will contribute to the prevention of organic fouling in various applications of NF
332 membranes.

333

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337

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425 (1986) 167–173.
- 426

427 Table 1 Typical compounds obtained from the pyrolysis of Wako humic acid (WHA), Aldrich
 428 humic acid (AHA), and organic colloids in groundwater condensed using the
 429 p(MPC-co-AEMA)-coated NF membrane under optimized hydrodynamic conditions (at 1500
 430 rpm and 0.75 MPa).

Peak No.	Compound	Group
1	Toluene	Aromatic
2	Octene	Aliphatic carbon
3	Ethylbenzene	Aromatic
4	Xylene	Aromatic
5	Styrene	Aromatic
6	Nonene	Aliphatic carbon
7	Phenol and Benzonitrile	Hydroxy benzene and N-containing
8	Decene and C ₃ -Alkylbenzene	Aliphatic carbon and aromatic
9	Undecene	Aliphatic carbon
10	Naphthalene	Aromatic
11	Dodecene	Aliphatic carbon
12	Tridecene	Aliphatic carbon
13	Methylnaphthalene	Aromatic
14	Biphenyl	Aromatic
15	Tetradecene	Aliphatic carbon
16	Pentadecene	Aliphatic carbon
17	Fluorene	Aromatic
18	Hexadecene	Aliphatic carbon
19	Heptadecene	Aliphatic carbon
20	Phenanthrene	Aromatic
21	Octadecene	Aliphatic carbon
22	Nonadecene	Aliphatic carbon
23	Methylphenanthrene and Methylantracene	Aromatic
24	Eicosene	Aliphatic carbon
25	Heneicosene	Aliphatic carbon
26	Docosene	Aliphatic carbon
27	Tricosene	Aliphatic carbon
28	Tetracosene	Aliphatic carbon
29	Pentacosene	Aliphatic carbon
30	Hexacosene	Aliphatic carbon
31	Heptacosene	Aliphatic carbon
32	Octacosene	Aliphatic carbon

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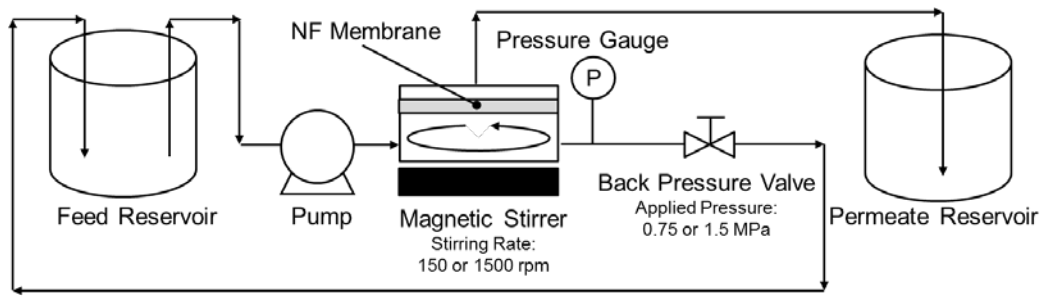


Fig. 1. Schematic of the cross-flow condensation apparatus.

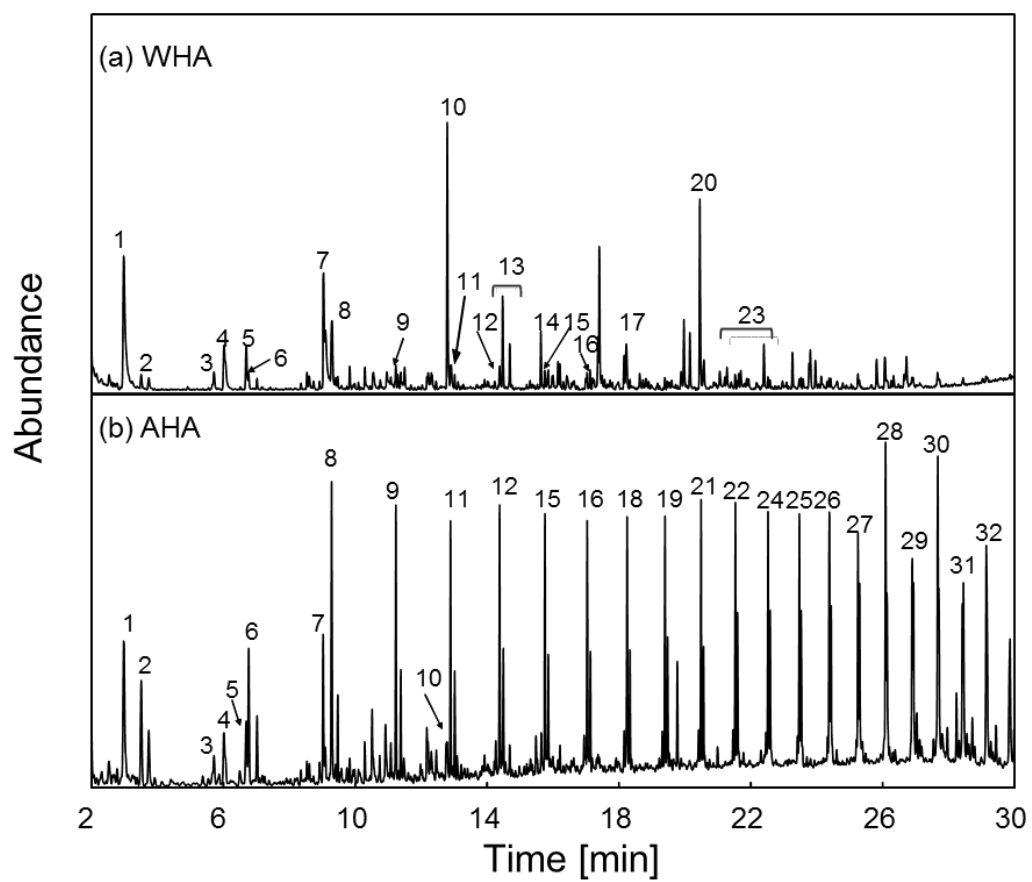


Fig. 2. Py-GC/MS pyrograms of (a) WHA and (b) AHA. Table 1 shows the peak numbers.

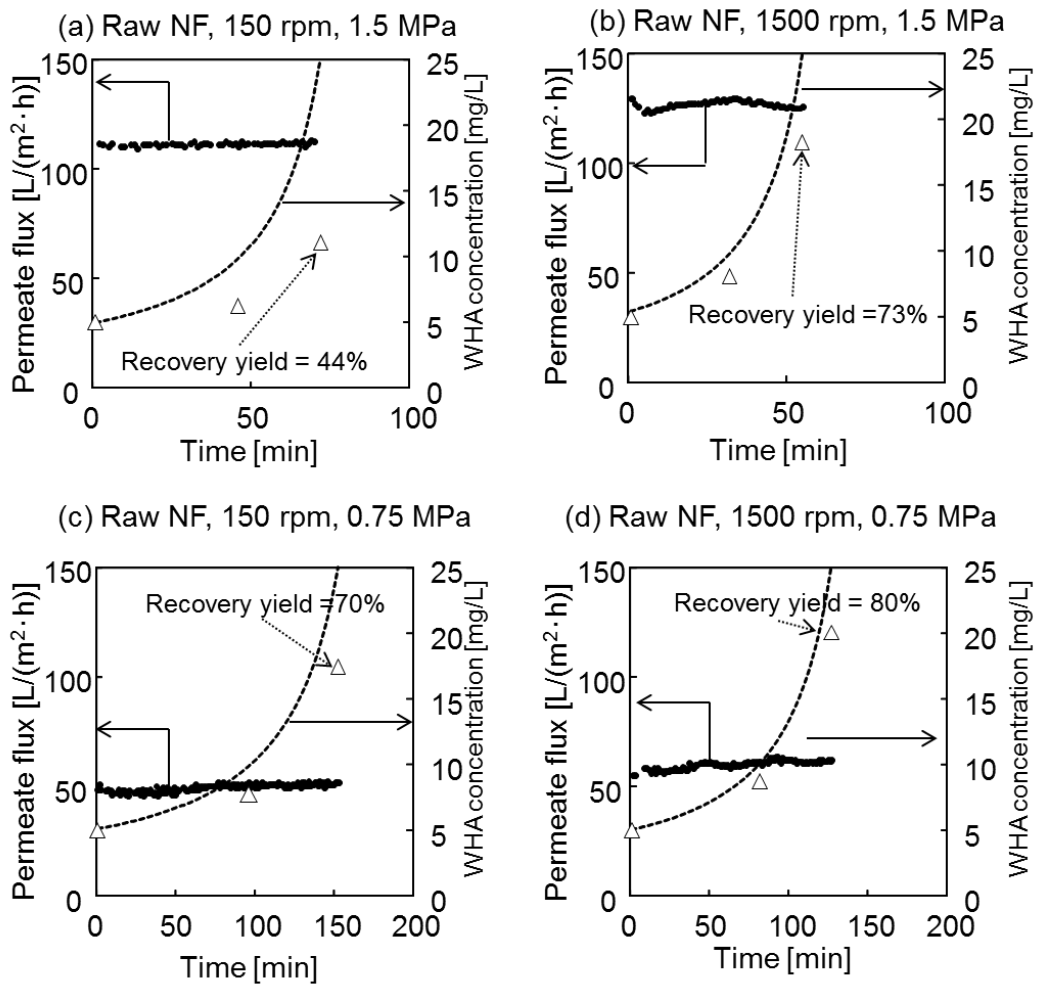


Fig. 3. Time courses of the permeate flux and WHA concentrations in the condensed solutions from the condensation tests of WHA using (a) raw NF membrane under the original hydrodynamic conditions (at 150 rpm and 1.5 MPa), (b) raw NF membrane at 1500 rpm and 1.5 MPa, (c) raw NF membrane at 150 rpm and 0.75 MPa, (d) raw NF membrane under the optimized hydrodynamic conditions (at 1500 rpm and 0.75 MPa). Closed circles and open triangles indicate permeate flux and WHA concentration, respectively. Dotted line indicates the theoretical concentrations of WHA assuming 100% recovery yield.

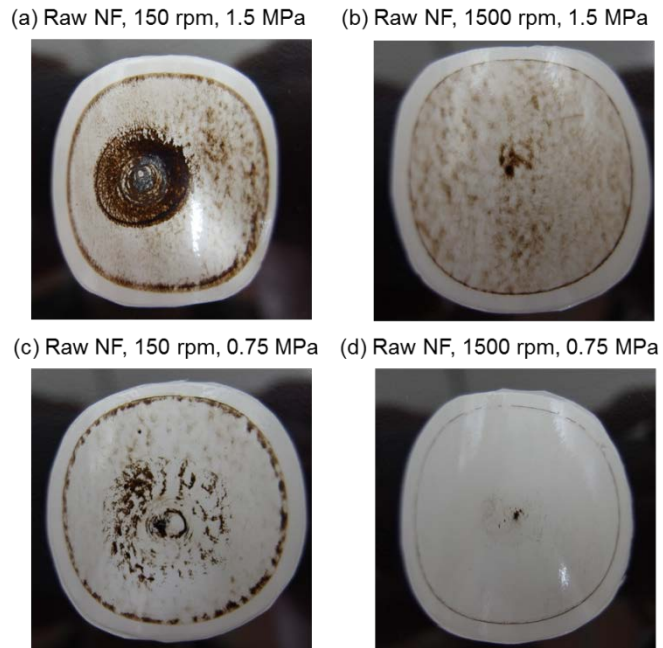


Fig. 4. Surface views of the membranes after the 5-fold condensation tests of WHA using (a) raw NF membrane under the original hydrodynamic condition (at 150 rpm and 1.5 MPa), (b) raw NF membrane at 1500 rpm and 1.5 MPa, (c) raw NF membrane at 150 rpm and 0.75 MPa, (d) raw NF membrane under the optimized hydrodynamic conditions (at 1500 rpm and 0.75 MPa).

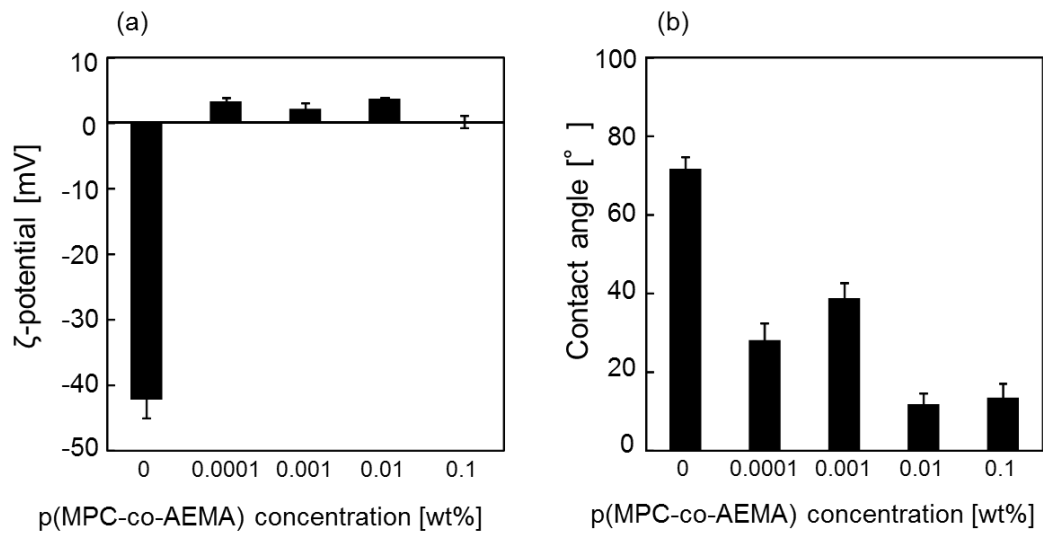


Fig. 5. ζ -potentials (a) and water contact angles (b) of the raw NF membrane and coated NF membranes with different concentrations of p(MPC-co-AEMA).

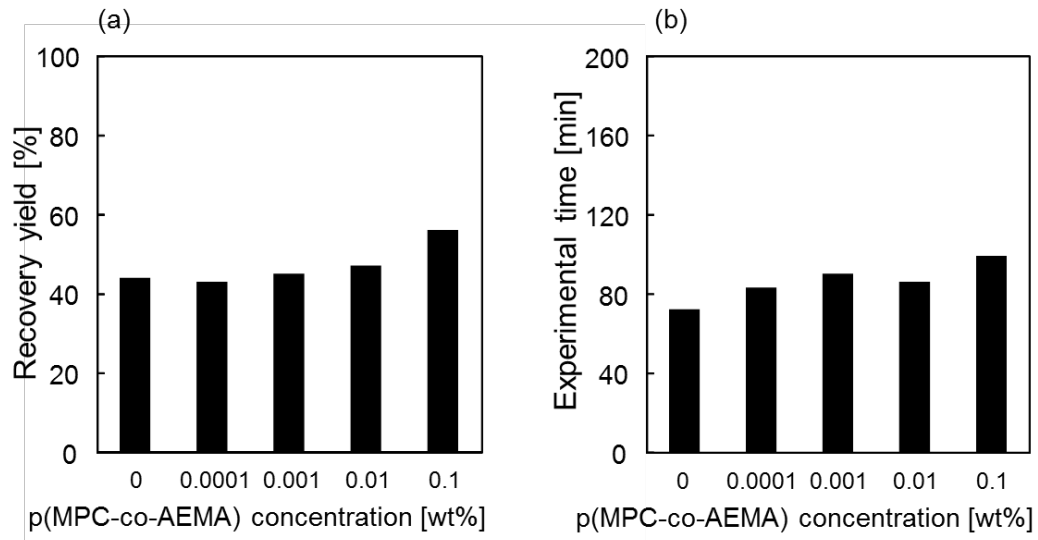


Fig. 6. Recovery yields (a) and experimental times (b) of the 5-fold concentration test of the humic acid at 1.5 MPa and 150 rpm using the raw NF membrane and coated NF membranes with different concentrations of p(MPC-co-AEMA).

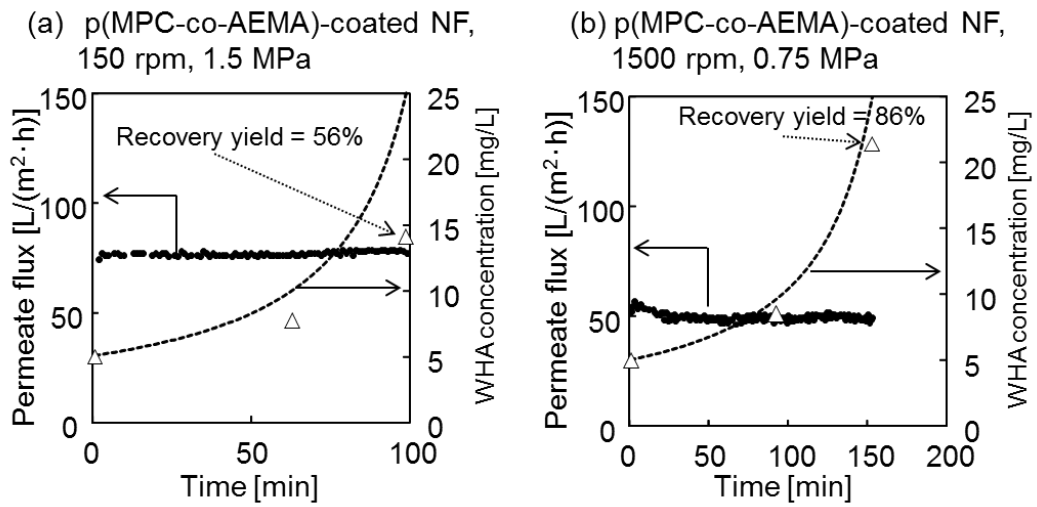


Fig. 7. Time courses of the permeate flux and WHA concentrations in the condensed solutions from the condensation tests of WHA using (a) p(MPC-co-AEMA)-coated NF membrane under the original hydrodynamic conditions (at 150 rpm and 1.5 MPa), (b) p(MPC-co-AEMA)-coated NF membrane under the optimized hydrodynamic conditions (at 1500 rpm and 0.75 MPa). Closed circles and open triangles indicate permeate flux and WHA concentration, respectively. Dotted line indicates the theoretical concentrations of WHA assuming 100% recovery yield.

(a) p(MPC-co-AEMA)-coated NF, 150 rpm, 1.5 MPa (b) p(MPC-co-AEMA)-coated NF, 1500 rpm, 0.75 MPa



Fig. 8. Surface views of the membranes after the 5-fold condensation tests of WHA using (a) p(MPC-co-AEMA)-coated NF membrane under the original hydrodynamic condition (at 150 rpm and 1.5 MPa), (b) p(MPC-co-AEMA)-coated NF membrane under optimized hydrodynamic conditions (at 1500 rpm and 0.75 MPa).

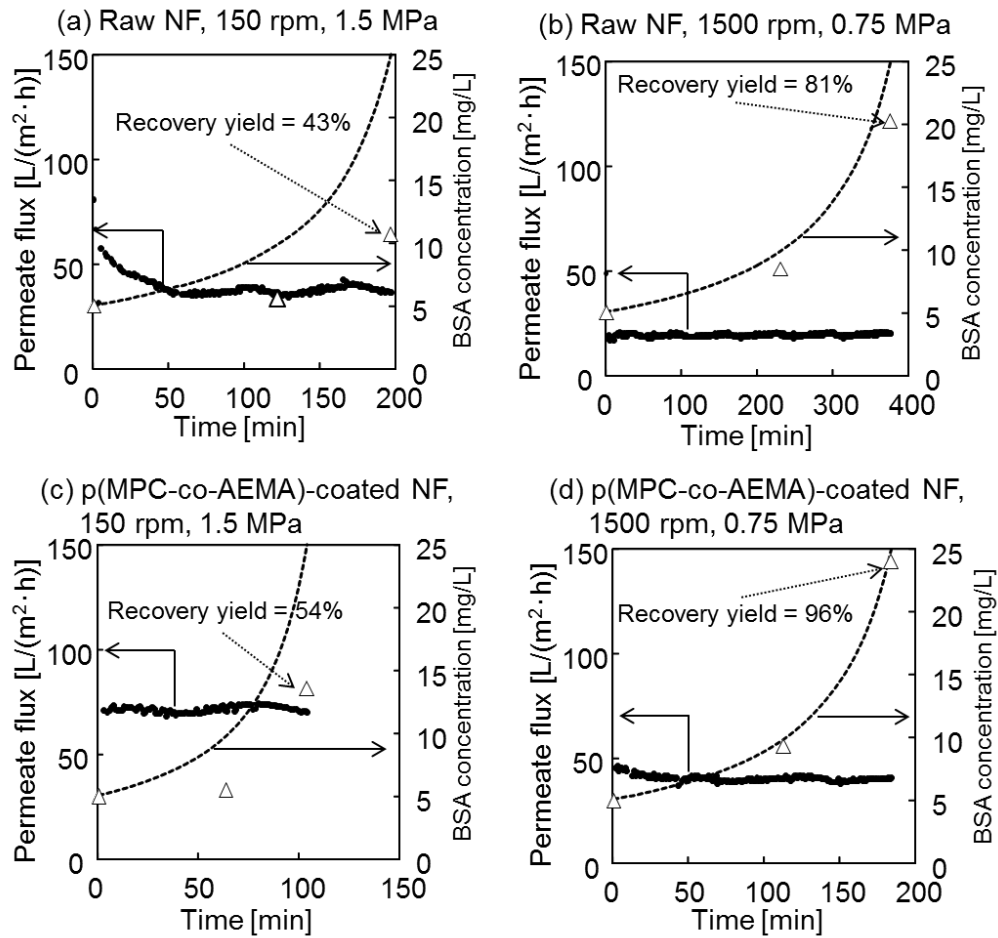


Fig. 9. Time courses of the permeate flux and BSA concentrations in the condensed solutions from the condensation tests of the BSA using (a) raw NF membrane under the original hydrodynamic condition (at 150 rpm and 1.5 MPa), (b) raw NF membrane under the optimized hydrodynamic conditions (at 1500 rpm and 0.75 MPa), (c) p(MPC-co-AEMA)-coated NF membrane under the original hydrodynamic condition (at 150 rpm and 1.5 MPa), (d) p(MPC-co-AEMA)-coated NF membrane under the optimized hydrodynamic conditions (at 1500 rpm and 0.75 MPa). Closed circles and open triangles indicate permeate flux and BSA concentration, respectively. Dotted line indicates the theoretical concentrations of BSA assuming 100% recovery yield.

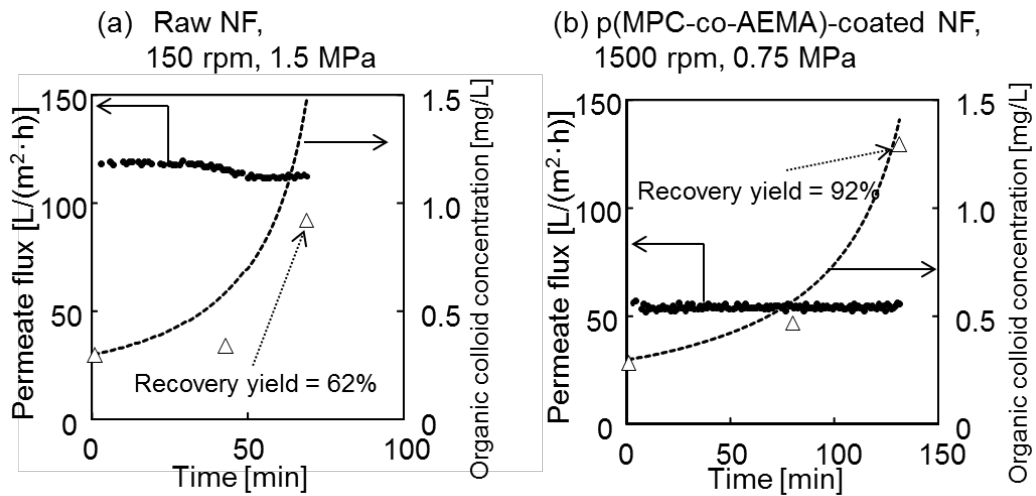


Fig. 10. Time courses of the permeate flux and organic colloid concentration of the condensed solutions from the condensation test of the groundwater using (a) raw NF membrane under the original hydrodynamic condition (at 150 rpm and 1.5 MPa), (b) p(MPC-co-AEMA)-coated NF membrane under the optimized hydrodynamic conditions (at 1500 rpm and 0.75 MPa). The concentration of organic colloids was calculated under the assumption that all organic colloids were WHA. Closed circles and open triangles indicate permeate flux and organic colloid concentration, respectively. Dotted line indicates theoretical concentrations of organic colloids assuming 100% recovery yield.

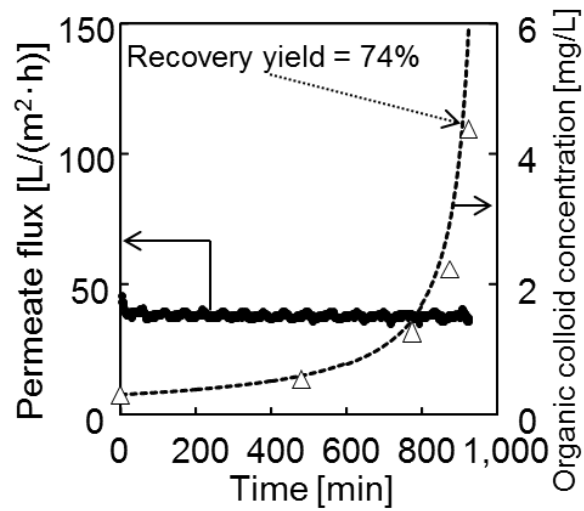


Fig. 11. Time courses of the permeate flux and organic colloid concentration in the condensed solutions from the condensation tests of the groundwater using the p(MPC-co-AEMA)-coated NF membrane under optimized hydrodynamic conditions (at 1500 rpm and 0.75 MPa). The groundwater was condensed from 500 mL to 25 mL. The concentration of organic colloids was calculated under the assumption that all organic colloids were WHA. Closed circles and open triangles indicate permeate flux and organic colloid concentration, respectively. Dotted line indicates theoretical concentrations of organic colloids assuming 100% recovery yield.

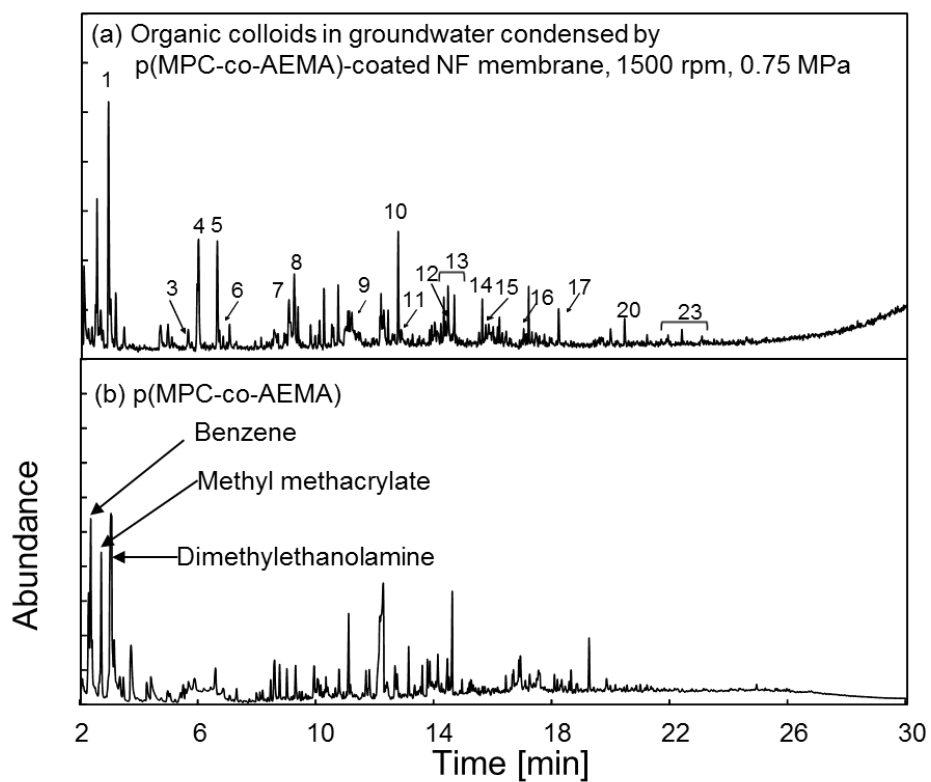


Fig. 12. Py-GC/MS pyrograms of (a) organic colloids in groundwater condensed by the p(MPC-co-AEMA)-coated NF membrane under optimized hydrodynamic conditions (at 1500 rpm and 0.75 MPa), (b) p(MPC-co-AEMA). Table 1 shows the peak numbers.